

Rapid Blood Pathogen Identification Panel

What is it?

The Methodist Microbiology Laboratory has introduced an FDA approved test called the Bloodstream Infection Test (performed by the Verigene® Gram-Positive or Gram-Negative Nucleic Acid Test). This is a qualitative, multiplexed *in vitro* diagnostic test which identifies genus, species and genetic antimicrobial resistance determinants for a broad panel of gram-positive *or* gram-negative bacteria (listed in **Table 1**) directly from positive blood culture bottles, allowing rapid identification of pathogens and earlier transition to most appropriate therapy. Information on which species are detected by the genus specific only assays can be found in **Table 4**.

Table 1: List of Pathogens Detected

Gram-positive bacteria	Gram-negative bacteria	Resistance Gene
<i>Staphylococcus</i> genus	<i>Escherichia coli</i>	mecA = methicillin (nafcillin) resistance
<i>Staphylococcus aureus</i>	<i>Klebsiella pneumoniae</i>	vanA = vancomycin resistance
<i>Staphylococcus epidermidis</i>	<i>Klebsiella oxytoca</i>	vanB = vancomycin resistance
<i>Staphylococcus lugdunensis</i>	<i>Pseudomonas aeruginosa</i>	CTX-M= ESBL
<i>Streptococcus</i> genus	<i>Serratia marcescens</i>	IMP = carbapenemase
<i>Streptococcus anginosus</i> Group	<i>Acinetobacter</i> genus	KPC = carbapenemase
<i>Streptococcus agalactiae</i>	<i>Citrobacter</i> genus	NDM = carbapenemase
<i>Streptococcus pneumoniae</i>	<i>Enterobacter</i> genus	OXA = carbapenemase
<i>Streptococcus pyogenes</i>	<i>Proteus</i> genus	VIM = carbapenemase
<i>Enterococcus faecalis</i>		
<i>Enterococcus faecium</i>		
<i>Listeria</i> genus		

How does it work?

When a blood culture bottle is identified as positive by the automated instrument, a microbiology technologist will perform a Gram stain on blood from the bottle. If the Gram stain shows a Gram-positive organism, the Gram-Positive Nucleic Acid Test will be performed and if the Gram stain shows a Gram-negative organism, the Gram-Negative Nucleic Acid Test will be performed. The results of the Bloodstream Infection Test will typically be available in Cerner within 3 hours of notification of a positive blood culture.

A list of recommended therapeutic decisions and treatment choices based on the Verigene® data are outlined in **Table 2 and Table 3**. These recommendations are based on an analysis of the Methodist Hospital antibiogram – please refer to the antibiogram for the most current susceptibility data from our lab.

Why is it helpful?

Previous studies have shown that rapid pathogen identification can result in earlier time to active therapy, earlier time to most appropriate therapy, shorter hospital stay, and improved clinical outcomes. The utility and cost-effectiveness of such testing is dependent on clinicians reacting to the data in real time. **It is strongly recommended that the Verigene results be utilized for therapy decisions at the time they are available.**

Certain infections are frequently polymicrobial in nature and the isolation of a single pathogen from the blood culture, while allowing narrowing of therapy, should not result in over-narrowing. An example of this would be a complicated intra-abdominal infection. This infection often involves anaerobes and therapy active against these should generally be included until definitive cultures of the site of infection have returned.

Final susceptibilities will be available in 24-72 hours and should always be reviewed to determine if therapy adjustments need to be made.

Table 2: Gram Positive Pathogen Detected and Recommended Therapy Adult Dosing

*For all gram positive bacteremia - If endocarditis is concern, consult ID for consideration of gentamicin

Pathogen Detected	Preferred Therapy	Comments
<i>Staphylococcus</i> genus with negative <i>S. aureus</i> PCR, including <i>S. epidermidis</i> and excluding <i>S. lugdunensis</i> <u>Blood Culture result:</u> 1 of 2 BCX positive	Consider withholding or discontinuing therapy as likely contaminant	In severely ill patients, consider starting/continuing therapy until more definitive results return.
2 of 2 BCX positive mecA negative	Nafcillin 2g q4h or 12g q24h	Cefazolin 2g q8h is an alternative
mecA positive	Vancomycin 15 mg/kg q12h	Pharmacokinetic dosing per pharmacy
<i>Staphylococcus lugdunensis</i>	Nafcillin 2g q4h or 12g q24h	<i>S. lugdunensis</i> is a virulent coagulase-negative staphylococcus that behaves like <i>S. aureus</i> and should be presumed to be pathogenic.

<p><i>Staphylococcus aureus</i> mecA negative = MSSA</p> <p>mecA positive = MRSA</p>	<p>Nafcillin 2g q4h or 12g q24h</p> <p>Vancomycin 15 mg/kg IV q12h</p>	<p>Cefazolin 2g q8h is an alternative</p> <p>Pharmacokinetic dosing per pharmacy</p>
<p><i>Streptococcus</i> genus <u>Blood Culture result:</u> 1 of 2 BCX positive</p> <p>2 of 2 BCX positive</p>	<p>Consider withholding or discontinuing therapy as likely contaminant</p> <p>Ceftriaxone 2g q24h</p>	<p>In severely ill patients consider starting/continuing therapy until more definitive results return.</p>
<p><i>Streptococcus anginosus</i> Group</p>	<p>Penicillin G 3-4 million units q4h or ceftriaxone 2g IV q24 h</p>	<p>Sub-group of <i>viridans group streptococci</i> with a propensity for abscess formation.</p>
<p><i>Streptococcus agalactiae</i> (Group B Strep)</p>	<p>Penicillin G 3-4 million units q4h or ampicillin 2g q4h or cefazolin 2g q8h</p>	<p>β-hemolytic strep are routinely susceptible to penicillin</p>
<p><i>Streptococcus pneumoniae</i> <u>Source of Infection:</u> Uncomplicated pneumonia</p> <p>CNS infection</p>	<p>Ceftriaxone 2g q 24h</p> <p>Ceftriaxone 2 g q 12h AND vancomycin 15 mg/kg q 12h</p>	<p>Levofloxacin 750mg Q24h is an alternative</p> <p>Continue vancomycin until susceptibilities return</p>
<p><i>Streptococcus pyogenes</i> (Group A Strep)</p>	<p>Penicillin G 3-4 million units q 4h or ampicillin 2g q4h or cefazolin 2g Q8h</p>	<p>β-hemolytic strep are routinely susceptible to penicillin</p>
<p><i>Enterococcus faecalis</i> or <i>Enterococcus faecium</i> van A/B negative</p> <p>van A/B positive = VRE</p>	<p>Ampicillin 2g IV q 4h</p> <p>Linezolid 600 mg q 12h or daptomycin 8 mg/kg IV q 24h</p>	<p>Alternate: Vancomycin 15 mg/kg IV q 12h</p>
<p><i>Listeria</i> genus</p>	<p>Nafcillin 2g Q4h or 12g q24h</p>	<p>TMP/SMX in patients with severe beta-lactam allergy</p>

Table 3: Gram Negative Pathogen Detected and Recommended Therapy

*For all gram negative bacteremia - increase ceftriaxone to 2g Q12h for CNS indication;
increase meropenem to 2g Q8h for CNS indication

Pathogen Detected	Preferred Therapy	Comments
<i>Escherichia coli</i>	<u>Community onset:</u> Ceftriaxone 2g q24h <u>Nosocomial onset:</u> Piperacillin/tazobactam 4.5 g q8h	
<i>Klebsiella pneumoniae</i>	<u>Community onset:</u> Ceftriaxone 2g q24h <u>Nosocomial onset:</u> Piperacillin/tazobactam 4.5 g q8h	
<i>Klebsiella oxytoca</i>	<u>Community onset:</u> Cefepime 1g q6h <u>Nosocomial onset:</u> Ertapenem 1g q24h	
<i>Pseudomonas aeruginosa</i>	Piperacillin/tazobactam 4.5g q8h OR cefepime 2g q8h	Consider addition of tobramycin in severely ill patients while awaiting susceptibility testing
<i>Serratia marcescens</i>	Ceftriaxone 2g q24h OR Cefepime 2g q8h	
<i>Acinetobacter</i> genus	Meropenem 1g q8h	No beta-lactam with >90% susceptibility Consider addition of amikacin in severely ill or non-responding
<i>Citrobacter</i> genus	Ceftriaxone 2g q24h Or piperacillin/tazobactam 4.5g q8h	
<i>Enterobacter</i> genus	Cefepime 2g q8h Or meropenem 1g q8h	
<i>Proteus</i> genus	Ceftriaxone 2g q24h	
CTX-M gene	Consult ID Meropenem 1g q8h	Marker for extended spectrum beta-lactamase (ESBL)
IMP, KPC, NDM, OXA or VIM genes	Consult ID Ceftazidime/avibactam (nonformulary) + Colistin +/- tigecycline	Marker for carbapenem resistant <i>Enterobacteriaceae</i> (CRE)

Table 4: Pathogens Detected by Genus Specific Assays

Genus Specific Assay	Pathogens Detected	Pathogens That May Not Be Detected
<i>Staphylococcus</i> genus	<i>S. arlettae</i> <i>S. aureus</i> <i>S. auricularis</i> <i>S. capitis</i> <i>S. caprae</i> <i>S. chromogenes</i> <i>S. cohnii</i> <i>S. epidermidis</i> <i>S. haemolyticus</i> <i>S. hominis</i> <i>S. intermedius</i> <i>S. lugdunensis</i> <i>S. muscae</i> <i>S. pasteurii</i> <i>S. saccharolyticus</i> <i>S. saprophyticus</i> <i>S. schleferi</i> <i>S. sciuri</i> <i>S. simulans</i> <i>S. warneri</i> <i>S. xylosus</i>	<i>S. carnosus</i> <i>S. condimentii</i> <i>S. delphini</i> <i>S. equorum</i> <i>S. felis</i> <i>S. hyicus</i> <i>S. gallinarum</i> <i>S. kloosii</i> <i>S. lentus</i> <i>S. lutrae</i> <i>S. nepalensis</i> <i>S. pettenkoferi</i> <i>S. piscifermentans</i> <i>S. flueretii</i> <i>S. pseudointerdium</i> <i>S. simiae</i> <i>S. succinus</i>
<i>Streptococcus</i> genus	<i>S. agalactiae</i> <i>S. anginosus</i> <i>S. bovis</i> <i>S. constellatus</i> <i>S. dysgalactiae</i> <i>S. dysgalactiae</i> subsp. <i>equismilis</i> <i>S. equi</i> <i>S. equinus</i> <i>S. gallolyticus</i> <i>S. gallolyticus pasteurianus</i> <i>S. gordonii</i> <i>S. infantarius</i> subsp. <i>coli</i> <i>S. infantarius</i> subsp. <i>infantarius</i> <i>S. infantis</i> <i>S. intermedius</i> <i>S. mitis</i> <i>S. mitis/oralis</i>	<i>S. acidominimus</i> <i>S. canis</i> <i>S. criceti</i> <i>S. downei</i> <i>S. ferus</i> <i>S. ictaluri</i> <i>S. iniae</i> <i>S. lactarius</i> <i>S. lutetiensis</i> <i>S. macacae</i> <i>S. macedonius</i> <i>S. parauberis</i> <i>S. pasteurianus</i> <i>S. pluranimalium</i> <i>S. porcinus</i> <i>S. pseudopneumoniae</i> <i>S. pseudoporcinus</i> <i>S. ratti</i> <i>S. suis</i>

	<i>S. parasanguinis</i> <i>S. peroris</i> <i>S. pneumoniae</i> <i>S. pyogenes</i> <i>S. salivarius</i> <i>S. sanguinis</i> <i>S. thoralensis</i>	<i>S. urinalis</i> <i>S. cristatus</i> <i>S. australis</i> <i>S. sinesis</i> <i>S. orisratti</i> <i>S. oligofermentans</i> <i>S. massiliensis</i> <i>S. sobrinus</i> <i>S. vestibularis</i> <i>S. thermophiles</i> <i>S. alactolyticus</i> <i>S. uberis</i> <i>S. equismilis</i>
<i>Listeria</i> genus	<i>L. innocua</i> <i>L. ivanovii</i> <i>L. monocytogenes</i> <i>L. seeligeri</i> <i>L. welshimeri</i>	<i>L. marthii</i> <i>L. grayi</i>
<i>Acinetobacter</i> genus	<i>A. baumannii</i> <i>A. baylyi</i> <i>A. bereziniae</i> <i>A. calcoaceticus</i> <i>A. guillouiae</i> <i>A. haemolyticus</i> <i>A. johnsonii</i> <i>A. junii</i> <i>A. lwoffii</i> <i>A. radioresistens</i> <i>A. schindleri</i> <i>A. ursingii</i>	<i>A. tartarogenes</i>
<i>Enterobacter</i> genus	<i>E. aerogenes</i> <i>E. amnigenus</i> <i>E. asburiae</i> <i>E. cancerogenus</i> <i>E. cloacae</i> <i>E. hormaechei</i> <i>E. ludwigii</i> <i>E. nimipressuralis/E. orzae</i>	<i>E. gergoviae</i> <i>E. kobei</i> <i>E. pyrinus</i>
<i>Proteus</i> genus	<i>P. hauseri</i> <i>P. mirabilis</i> <i>P. myxofaciens</i> <i>P. penneri</i> <i>P. vulgaris</i>	